

POCKET-SIZED DEVICES - **FLUORPEN**

LIST OF REFERENCES

AJIGBOYE O. O., LU CH., MURCHIE E. H., ET AL. (2017). Altered gene expression by sedaxane increases PSII efficiency, photosynthesis and growth and improves tolerance to drought in wheat seedlings. *Pesticide Biochemistry and Physiology*. Volume 137. Pages 49-61.

DOI: 10.1016/j.pestbp.2016.09.008.

<http://www.sciencedirect.com/science/article/pii/S0048357516301109>

CHEKANOV K., SCHASTNAYA E., SOLOVCHENKO A., ET AL. (2017). Effects of CO₂ enrichment on primary photochemistry, growth and astaxanthin accumulation in the chlorophyte *Haematococcus pluvialis*. *Journal of Photochemistry and Photobiology B: Biology*. Volume 171.

DOI 10.1016/j.jphotobiol.2017.04.028.

<https://www.ncbi.nlm.nih.gov/pubmed/28475936>

DUARTE B., PEDRO S., MARQUES J. C., ET AL. (2017). *Zostera noltii* development probing using chlorophyll a transient analysis (JIP-test) under field conditions: Integrating physiological insights into a photochemical stress index. *Ecological Indicators*. Volume 76.

DOI: 10.1016/j.ecolind.2017.01.023.

<http://www.sciencedirect.com/science/article/pii/S1470160X17300298>

HERNÁNDEZ-CLEMENTE R., NORTH P.R.J., HORNERO A., ET AL. (2017). Assessing the effects of forest health on sun-induced chlorophyll fluorescence using the FluorFLIGHT 3-D radiative transfer model to account for forest structure, *Remote Sensing of Environment*,. Volume 193. Pages 165-179.

DOI: 10.1016/j.rse.2017.02.012.

<http://www.sciencedirect.com/science/article/pii/S0034425717300718>

LEE M. W., HUFFAKER A., CRIPPEN D., ET AL. (2017). Plant Elicitor Peptides Promote Plant Defenses against Nematodes in Soybean. *Molecular Plant Pathology*.

DOI: 10.1111/mpp.12570

<https://www.ncbi.nlm.nih.gov/pubmed/28600875>

MARTEL A. B. AND QADERI M. M. (2017). Light quality and quantity regulate aerobic methane emissions from plants. *Physiol Plantarum*. Volume 159.

DOI:10.1111/ppl.12514

<http://onlinelibrary.wiley.com/doi/10.1111/ppl.12514/abstract>

PARADISO R., ARENA C., DE MICCO V., ET AL. (2017). Changes in Leaf Anatomical Traits Enhanced Photosynthetic Activity of Soybean Grown in Hydroponics with Plant Growth-Promoting Microorganisms. *Frontiers in Plant Science*. Volume 8.

DOI: 10.3389/fpls.2017.00674

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5418343/>

PTUSHENKO V.V., PTUSHENKO O.S., SAMOILOVA O.P., ET AL. (2017). *The analysis of photoprotection and apparent non-photochemical quenching of chlorophyll fluorescence in Tradescantia leaves based on the rate of irradiance-induced changes in optical transparency. Biochemistry (Moscow). Volume 82.*

DOI: 10.1134/S0006297917010072

<https://link.springer.com/article/10.1134/S0006297917010072>

PUGLIELLI G., REDONDO-GÓMEZ S., GRATANI L., ET AL. (2017). *Highlighting the differential role of leaf paraheliotropism in two Mediterranean Cistus species under drought stress and well-watered conditions. Journal of Plant Physiology. Volume 213.*

DOI: 10.1016/j.jplph.2017.02.015.

<http://www.sciencedirect.com/science/article/pii/S0176161717300895>

PUSHKAREVA E., KVÍDEROVÁ J., ŠIMEK M., ET AL. (2017). *Nitrogen fixation and diurnal changes of photosynthetic activity in Arctic soil crusts at different development stage. European Journal of Soil Biology. Volume 79. Pages 21-30*

DOI: 10.1016/j.ejsobi.2017.02.002.

<http://www.sciencedirect.com/science/article/pii/S1164556317300742>

SELVARAJ, M.G., ISHIZAKI, T., VALENCIA, M., ET AL. (2017). *Overexpression of an Arabidopsis thaliana galactinol synthase gene improves drought tolerance in transgenic rice and increased grain yield in the field. Plant Biotechnol. J.*

DOI: 10.1111/pbi.12731

<http://onlinelibrary.wiley.com/doi/10.1111/pbi.12731/abstract>

SINGH M., KUSHWAHA K. B., SINGH, S., ET AL. (2017). *Sulphur alters chromium (VI) toxicity in Solanum melongena seedlings: Role of sulphur assimilation and sulphur-containing antioxidants. Plant Physiology and Biochemistry. Volume 112.*

DOI: 10.1016/j.plaphy.2016.12.024.

<http://www.sciencedirect.com/science/article/pii/S0981942816304910>

SINGH S. & PRASAD S. M. (2017) *Effects of 28-homobrassinoloid on key physiological attributes of Solanum lycopersicum seedlings under cadmium stress: Photosynthesis and nitrogen metabolism. Plant Growth Regul J. Volume 82.*

DOI:10.1007/s10725-017-0248-5

<https://link.springer.com/article/10.1007/s10725-017-0248-5>

TRIPATHI D. K., MISHRA R. K., SINGH S., ET AL. (2017). *Nitric Oxide Ameliorates Zinc Oxide Nanoparticles Phytotoxicity in Wheat Seedlings: Implication of the Ascorbate–Glutathione Cycle. Frontiers in Plant Science. Volume 8.*

DOI: 10.3389/fpls.2017.00001

<http://doi.org/10.3389/fpls.2017.00001>

TRIPATHI D. K., SINGH S., SINGH S., ET AL. (2017). *Nitric oxide alleviates silver nanoparticles (AgNps)-induced phytotoxicity in Pisum sativum seedlings. Plant Physiology and Biochemistry. Volume 110.*

DOI: 10.1016/j.plaphy.2016.06.015.

<http://www.sciencedirect.com/science/article/pii/S0981942816302364>

TRIPATHI D. K., SINGH S., SINGH V. P., ET AL. (2017). Silicon nanoparticles more effectively alleviated UV-B stress than silicon in wheat (*Triticum aestivum*) seedlings, *Plant Physiology and Biochemistry*. Volume 110.

DOI: 10.1016/j.plaphy.2016.06.026.

<http://www.sciencedirect.com/science/article/pii/S0981942816302479>

ZORIN B., PAL-NATH D., LUKYANOV A., ET AL. (2017). Arachidonic acid is important for efficient use of light by the microalga *lobo-sphaera incisa* under chilling stress. *Biochimica et Biophysica Acta - Molecular and Cell Biology of Lipids*.

DOI 10.1016/j.bbalip.2017.04.008

<https://www.ncbi.nlm.nih.gov/pubmed/28504210>

AHMED S., ARIYARATNE M., PATEL J., ET AL. (2016). Altered expression of polyamine transporters reveals a role for spermidine in the timing of flowering and other developmental response pathways., *Plant Science*.

DOI: 10.1016/j.plantsci.2016.12.002

<http://www.sciencedirect.com/science/article/pii/S0168945216303740>

AJIGBOYE O. O., BOUSQUET L., MURCHIE E. H. ET AL. (2016). Chlorophyll fluorescence parameters allow the rapid detection and differentiation of plant responses in three different wheat pathosystems. *Functional Plant Biology*. Volume 43. Pages 356–369.

DOI: 10.1071/FP15280

<http://www.publish.csiro.au/paper/FP15280.htm>

AJIGBOYE O.O., LU C., MURCHIE E. H. ET AL. (2016) Altered gene expression by sedaxane increases PSII efficiency, photosynthesis and growth and improves tolerance to drought in wheat seedlings. *Pesticide Biochemistry and Physiology*.

DOI: 10.1016/j.pestbp.2016.09.008

<https://www.researchgate.net/publication/308891905> Altered gene expression by sedaxane increases P SII efficiency photosynthesis and growth and improves tolerance to drought in wheat seedlings

ATHAR H. R., AMBREEN S., JAVED M. ET AL. (2016) Influence of sub-lethal crude oil concentration on growth, water relations and photosynthetic capacity of maize (*Zea mays* L.) plants. *Environmental Science and Pollution Research*. Volume 23, Issue 18, Pages 18320–18331.

DOI: 10.1007/s11356-016-6976-7

<https://www.ncbi.nlm.nih.gov/pubmed/27278069>

BARÁNYIOVÁ I. AND KLEM K. (2016) Effect of application of growth regulators on the physiological and yield parameters of winter wheat under water deficit. *Plant, Soil and Environment*. Volume 62, No. 3, Pages 114–120.

DOI: 10.17221/778/2015-PSE

<http://www.agriculturejournals.cz/publicFiles/178110.pdf>

CHEKANOV K., LUKYANOV A., BOUSSIBA S. ET AL. (2016) Modulation of photosynthetic activity and photoprotection in *Haematococcus pluvialis* cells during their conversion into haematocysts and back. *Photosynthesis Research*. Volume 128, Issue 3, Pages 313–323.

DOI: 10.1007/s11120-016-0246-x

<https://www.ncbi.nlm.nih.gov/pubmed/27002330>

CHOI H. G., MOON B. Y. AND KANG N. J. (2016). Correlation between Strawberry (*Fragaria ananassa* Duch.) Productivity and Photosynthesis-Related Parameters under Various Growth Conditions. *Frontiers in Plant Science*. Volume 7.

DOI: 10.3389/fpls.2016.01607

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5080357/>

DUARTE B., CABRITA M. T., GAMEIRO C. ET AL. (2016) Disentangling the photochemical salinity tolerance in *Aster tripolium* L. : Connecting biophysical traits with changes in the fatty acid composition. *Plant biology*.

DOI: 10.1111/plb.12517

<https://www.ncbi.nlm.nih.gov/pubmed/27748562>

ESTEBAN R., ROYO B., URARTE E. ET AL. (2016) Both Free Indole-3-Acetic Acid and Photosynthetic Performance are Important Players in the Response of *Medicago truncatula* to Urea and Ammonium Nutrition Under Axenic Conditions. *Frontiers in Plant Science*. Volume 7.

DOI: 10.3389/fpls.2016.00140

<https://www.ncbi.nlm.nih.gov/pubmed/26909089>

FROSI G., BARROS V. A., OLIVEIRA M. T., ET AL. (2016). Symbiosis with AMF and leaf Pi supply increases water deficit tolerance of woody species from seasonal dry tropical forest. *Journal of Plant Physiology*. Volume 207.

DOI: 10.1016/j.jplph.2016.11.002.

<http://www.sciencedirect.com/science/article/pii/S0176161716302449>

HIDRI R., BAREA J.M., MAHMOUD O. MB. ET AL. (2016) Impact of microbial inoculation on biomass accumulation by *Sulla carnosa* provenances, and in regulating nutrition, physiological and antioxidant activities of this species under non-saline and saline conditions. *Journal of Plant Physiology*. Volume 201, Pages 28–41.

DOI: 10.1016/j.jplph.2016.06.013

<https://www.ncbi.nlm.nih.gov/pubmed/27393918>

KANECHI M., MAEKAWA A., NISHIDA Y. AND MIYASHITA, E. (2016). Effects of pulsed lighting based light-emitting diodes on the growth and photosynthesis of lettuce leaves. *Acta Hort.* Volume 1134.

DOI: 10.17660/ActaHortic.2016.1134.28

<https://doi.org/10.17660/ActaHortic.2016.1134.28>

LV DW., ZHU GR., ZHU D. ET AL. (2016) Proteomic and phosphoproteomic analysis reveals the response and defense mechanism in leaves of diploid wheat *T. monococcum* under salt stress and recovery. *Journal of Proteomics*. Volume 143, Pages 93–105.

DOI: 10.1016/j.jprot.2016.04.013

<https://www.ncbi.nlm.nih.gov/pubmed/27095598>

LÓPEZ-LÓPEZ M., CALDERÓN R., GONZÁLEZ-DUGO V., ET L. (2016). Early Detection and Quantification of Almond Red Leaf Blotch Using High-Resolution Hyperspectral and Thermal Imagery. *Remote Sens*. Volume 8.

DOI:10.3390/rs8040276

<http://www.mdpi.com/2072-4292/8/4/276/htm>

MALIK V.M., LOBO J.M., STEWART C. ET AL. (2016). Growth irradiance affects ureide accumulation and tolerance to photoinhibition in *Eutrema salsugineum* (*Thellungiella salsuginea*). *Photosynthetica*. Volume 54.

DOI:10.1007/s11099-015-0164-8

<http://link.springer.com/article/10.1007/s11099-015-0164-8>

MARTEL A. B. AND QADERI M. M. (2016), *Light quality and quantity regulate aerobic methane emissions from plants. Physiol Plantarum.*

DOI: 10.1111/ppl.12514

<http://onlinelibrary.wiley.com/doi/10.1111/ppl.12514/full>

MISHRA R. K. KUMAR J., SRIVASTAVA P. K., ET AL. (2016). *PSII photochemistry, oxidative damage and anti-oxidative enzymes in arsenate-stressed Oryza sativa L. seedlings. Volume 33.*

DOI: 10.1080/02757540.2016.1265516

<http://www.tandfonline.com/doi/citedby/10.1080/02757540.2016.1265516?scroll=top&needAccess=true>

NAUŠ J., ŠMECKO S. AND ŠPUNDOVÁ M. (2016) *Chloroplast avoidance movement as a sensitive indicator of relative water content during leaf desiccation in the dark. Photosynthesis Research. Volume 129, Issue 2, Pages 217–225.*

DOI: 10.1007/s11120-016-0291-5

<http://link.springer.com/article/10.1007/s11120-016-0291-5>

OYIGA B. C., SHARMA R. C., SHEN J., (2016), *Identification and Characterization of Salt Tolerance of Wheat Germplasm Using a Multivariable Screening Approach. J Agro Crop Sci, 202: 472–485.*

DOI: 10.1111/jac.12178

<http://onlinelibrary.wiley.com/doi/10.1111/jac.12178/full>

PANEQUE M, DE LA ROSA J. M., FRANCO-NAVARRO J. D., ET AL. (2016). *Effect of biochar amendment on morphology, productivity and water relations of sunflower plants under non-irrigation conditions, CATENA, Volume 147.*

DOI: 10.1016/j.catena.2016.07.037.

<http://www.sciencedirect.com/science/article/pii/S034181621630296X>

PEDRANZANI H., RODRÍGUEZ-RIVERA M., GUTIÉRREZ M. ET AL. (2016) *Arbuscular mycorrhizal symbiosis regulates physiology and performance of Digitaria eriantha plants subjected to abiotic stresses by modulating antioxidant and jasmonate levels. Mycorrhiza. Volume 26, Issue 2, Pages 141–152.*

DOI: 10.1007/s00572-015-0653-4

<https://www.ncbi.nlm.nih.gov/pubmed/26184604>

PTUSHENKO V. V. AND SOLOVCHENKO, A. E. (2016). *Tolerance of the photosynthetic apparatus to acidification of the growth medium as a possible determinant of CO₂-tolerance of the symbiotic microalga DESMODESMUS sp. IPPAS-2014. Biochemistry Moscow. Volume 81.*

DOI:10.1134/S0006297916120142

<http://link.springer.com/article/10.1134%2FS0006297916120142>

RUIZ-LOZANO J. M., AROCA R., ZAMARREÑO Á. M. ET AL. (2016) *Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. Plant, Cell and Environment. Volume 39, Pages 441–452.*

DOI: 10.1111/pce.12631.

<https://www.ncbi.nlm.nih.gov/pubmed/26305264>

SOLOVCHENKO A., GORELOVA O., SELYAKH I., ET AL. (2016). *Nitrogen availability modulates CO₂ tolerance in a symbiotic chlorophyte. Algal Research. 2016. 16. 177-188.*

DOI 10.1016/j.algal.2016.03.002

<https://www.researchgate.net/publication/296486056> Nitrogen availability modulates CO₂ tolerance in a symbiotic chlorophyte

SRINIVASARAO CH., SHANKER A. K., KUNDU S. AND REDDY S. (2016) Chlorophyll fluorescence induction kinetics and yield responses in rainfed crops with variable potassium nutrition in K deficient semi-arid alfisols. *Journal of Photochemistry and Photobiology B: Biology*. Volume 160, Pages 86–95.

DOI: 10.1016/j.jphotobiol.2016.03.052

<http://www.sciencedirect.com/science/article/pii/S1011134415300798>

TIMM C. M., PELLETIER D. A., JAWDY S. S. ET AL. (2016) Two Poplar-Associated Bacterial Isolates Induce Additive Favorable Responses in a Constructed Plant-Microbiome System. *Frontiers in Plant Science*. Volume 7.

DOI: 10.3389/fpls.2016.00497

<https://www.ncbi.nlm.nih.gov/pubmed/27200001>

WATANABE T., ORIKASA T., SHONO H., ET AL. (2016). The influence of inhibit avoid water defect responses by heat pretreatment on hot air drying rate of spinach, *Journal of Food Engineering*, Volume 168.

DOI: 10.1016/j.jfoodeng.2015.07.014.

<http://www.sciencedirect.com/science/article/pii/S0260877415003131>

WEI J., YANG H., CAO H. AND TAN T. (2016) Using polyaspartic acid hydro-gel as water retaining agent and its effect on plants under drought stress. *Saudi Journal of Biological Sciences*. Volume 23, Pages 654–659.

DOI: 10.1016/j.sjbs.2015.08.016

<http://www.sciencedirect.com/science/article/pii/S1319562X15001941>

ZANDALINAS S. I., RIVERO R. M., MARTÍNEZ V., ET AL. (2016). Tolerance of citrus plants to the combination of high temperatures and drought is associated to the increase in transpiration modulated by a reduction in abscisic acid levels. *BMC Plant Biology* BMC. Volume 16.

DOI: 10.1186/s12870-016-0791-7

<https://bmcpantbiol.biomedcentral.com/articles/10.1186/s12870-016-0791-7>

ZARCO-TEJADA P.J., GONZÁLEZ-DUGO M.V. AND FERERES E. (2016) Seasonal stability of chlorophyll fluorescence quantified from airborne hyperspectral imagery as an indicator of net photosynthesis in the context of precision agriculture. *Remote Sensing of Environment*. Volume 179. Pages 89–103.

DOI: 10.1016/j.rse.2016.03.024

<http://www.sciencedirect.com/science/article/pii/S0034425716301183>

ANDERSON L. G., DUNN A. M., ROSEWARNE P. J. AND STEBBING P. D. (2015) Invaders in hot water: a simple decontamination method to prevent the accidental spread of aquatic invasive non-native species. *Biological Invasions*. Volume 17, Issue 8, Pages 2287–2297.

DOI: 10.1007/s10530-015-0875-6

<https://www.ncbi.nlm.nih.gov/pubmed/26316842>

ARMADA E., BAREA J.M., CASTILLO P. ET AL. (2015) Characterization and management of autochthonous bacterial strains from semiarid soils of Spain and their interactions with fermented agrowastes to improve drought tolerance in native shrub species. *Applied Soil Ecology*. Volume 96, Pages 306–318.

DOI: 10.1016/j.apsoil.2015.08.008

<http://www.sciencedirect.com/science/article/pii/S0929139315300615>

ARMADA E., AZCÓN R., LÓPEZ-CASTILLO O. M. ET AL. (2015) Autochthonous arbuscular mycorrhizal fungi and *Bacillus thuringiensis* from a degraded Mediterranean area can be used to improve physiological traits and performance of a plant of agronomic interest under drought conditions. *Plant Physiology and Biochemistry*. Volume 90, Pages 64–74.

DOI: 10.1016/j.plaphy.2015.03.004

<https://www.ncbi.nlm.nih.gov/pubmed/25813343>

BARTÁK M., TRNKOVÁ K., HANSEN E. S. ET AL. (2015) Effect of dehydration on spectral reflectance and photosynthetic efficiency in *Umbilicaria arctica* and *U. hyperborean*. *Biologia Plantarum*. Volume 59, Issue 2, Pages 357–365.

DOI: 10.1007/s10535-015-0506-1

<http://link.springer.com/article/10.1007/s10535-015-0506-1>

BÁRZANA G., AROCA R. AND RUIZ-LOZANO J. M. (2015) Localized and non-localized effects of arbuscular mycorrhizal symbiosis on accumulation of osmolytes and aquaporins and on antioxidant systems in maize plants subjected to total or partial root drying. *Plant, Cell and Environment*. Volume 38, Issue 8, Pages 1613–1627.

DOI: 10.1111/pce.12507

<http://onlinelibrary.wiley.com/doi/10.1111/pce.12507/abstract>

DUARTE B., GOESSLING J.W., MARQUES J.C. AND CAÇADOR I. (2015) Ecophysiological constraints of *Aster tripolium* under extreme thermal events impacts: Merging biophysical, biochemical and genetic insights. *Plant Physiology and Biochemistry*. Volume 97, Pages 217–228.

DOI: 10.1016/j.plaphy.2015.10.015

<https://www.ncbi.nlm.nih.gov/pubmed/26485432>

DYAKOV, M. Y., INSAROVA, I. D., KHARABADZE, D. E. ET AL. (2015). Influence of extreme ambient temperatures and anaerobic conditions on *Peltigera aphthosa* (L.) Willd. viability". *Life sciences in space research*. Volume 7.

DOI:10.1016/j.lssr.2015.10.002

<http://www.sciencedirect.com/science/article/pii/S2214552415000838>

FESENKO I. A., ARAPIDI G. P., SKRIPNIKOV A. Y., ET AL. (2015). Specific pools of endogenous peptides are present in gametophore, protonema, and protoplast cells of the moss *Physcomitrella patens*. *Pesticide Biochemistry and Physiology*. Volume 15, Pages 1-16.

DOI 10.1186/s12870-015-0468-7

<http://link.springer.com/article/10.1186%2Fs12870-015-0468-7>

HUMPLÍK J. F., LAZÁR D., FÜRST T. ET AL. (2015). Automated integrative high-throughput phenotyping of plant shoots: a case study of the cold-tolerance of pea (*Pisum sativum*L.). *Plant Methods*. Volume 11, Pages 1-11.

DOI 10.1186/s13007-015-0063-9

<http://link.springer.com/article/10.1186%2Fs13007-015-0063-9>

JIMÉNEZ J. D. L. C., CARDOSO J. A., DOMINGUEZ M. ET AL. (2015) Morpho-anatomical traits of root and non-enzymatic antioxidant system of leaf tissue contribute to waterlogging tolerance in Brachiaria grasses. *Grassland Science*. Volume 61, Pages 243–252.

DOI:10.1111/grs.12095

<http://onlinelibrary.wiley.com/doi/10.1111/grs.12095/abstract>

KHALID A., ATHAR H., ZAFAR Z. U. ET AL. (2015) Photosynthetic capacity of canola (*Brassica napus* L.) plants as affected by glycinebetaine under salt stress. *Journal of Applied Botany and Food Quality*. Volume 88, Pages 78- 86.

DOI:10.5073/JABFQ.2015.088.011

<http://pub.jki.bund.de/index.php/JABFQ/article/view/3074>

KOSOVÁ K., VÍTÁMVÁS P., HLAVÁČKOVÁ I. ET AL. (2015) Responses of two barley cultivars differing in their salt tolerance to moderate and high salinities and subsequent recovery. *Biologia Plantarum*. Volume 59, Pages 106-114.

DOI: 10.1007/s10535-014-0465-y

<http://link.springer.com/article/10.1007/s10535-014-0465-y>

MARQUEZ-GARCIA B., SHAW D., COOPER J. W. ET AL. (2015) Redox markers for drought-induced nodule senescence, a process occurring after drought-induced senescence of the lowest leaves in soybean (*Glycine max*). *Annals of Botany*. Volume 116, Pages 497–510.

DOI: 10.1093/aob/mcv030

<http://aob.oxfordjournals.org/content/early/2015/04/06/aob.mcv030.full>

OREKHOVA D. I., YAKOVLEVAB O. V., GORYACHEVB S. N., ET AL. (2015). The Use of Parameters of Chlorophyll a Fluorescence Induction to Evaluate the State of Plants under Anthropogenic Load. *Biophysics*. Volume 60, Pages 330–336.

DOI: 10.1134/S0006350915020128

<http://link.springer.com/article/10.1134%2FS0006350915020128#/page-1>

PTUSHENKO V. V., AVERCHEVA O. V., BASSARSKAYA E. M. ET AL. (2015) Possible reasons of a decline in growth of Chinese cabbage under a combined narrowband red and blue light in comparison with illumination by high-pressure sodium lamp. *Scientia Horticulturae*. Volume 194, Pages 267-277.

DOI:10.1016/j.scienta.2015.08.021

<http://www.sciencedirect.com/science/article/pii/S0304423815301400>

SOLOVCHENKO A., GORELOVA O., SELYAKH I., ET AL. (2015). A novel CO₂-tolerant symbiotic *Desmododesmus* (*Chlorophyceae*, *Desmodesmaceae*): acclimation to and performance at a high carbon dioxide level. *Algal Research*. Volume 11.

DOI 10.1016/j.algal.2015.04.011

<http://www.bashanfoundation.org/alexei/2015.-Solovchenko-AR.pdf>

ŠEBELA D., QUIÑONES C., OLEJNÍČKOVÁ J. AND JAGADISH K.S.V. (2015) Temporal chlorophyll fluorescence signals to track changes in optical properties of maturing rice panicles exposed to high night temperature. *Field Crops Research*. Volume 177, Pages 75–85.

DOI: 10.1016/j.fcr.2015.02.025

<http://www.sciencedirect.com/science/article/pii/S0378429015000775>

TRIPATHI D. K., SINGH V. P., PRASAD S. M. ET AL. (2015). Silicon-mediated alleviation of Cr(VI) toxicity in wheat seedlings as evidenced by chlorophyll fluorescence, laser induced breakdown spectroscopy and anatomical changes. *Ecotoxicology and Environmental Safety*, Volume 113, Pages 133-144.

DOI:10.1016/j.ecoenv.2014.09.029

<http://www.sciencedirect.com/science/article/pii/S0147651314004461>

WU X., TANG Y., LI C. ET AL. (2015) Chlorophyll Fluorescence and Yield Responses of Winter Wheat to Waterlogging at Different Growth Stages. *Plant Production Science*. Volume 18, Issue 3.

DOI: 10.1626/pps.18.284

https://www.jstage.jst.go.jp/article/pps/18/3/18_284/article

ZMIENKO A., GORALSKI M., SAMELAK-CZAJKA A. ET AL. (2015) Time course transcriptional profiling of senescing barley leaves. *Genomics Data*. Volume 4, Pages 78–81.

DOI: 10.1016/j.gdata.2015.03.006

<http://www.sciencedirect.com/science/article/pii/S2213596015000239>

AJIGBOYE O. O., MURCHIE E., RAY R. V. (2014). Foliar application of isopyrazam and epoxiconazole improves photosystem II efficiency, biomass and yield in winter wheat. *Pesticide Biochemistry and Physiology*. Volume 114, Pages 52–60.

DOI:10.1016/j.pestbp.2014.07.003

<http://www.sciencedirect.com/science/article/pii/S0048357514001278>

PTUSHENKO V. V., PTUSHENKO O. S. AND TIKHONOV A. N. (2014) Chlorophyll Fluorescence Induction, Chlorophyll Content, and Chromaticity Characteristics of Leaves as Indicators of Photosynthetic Apparatus Senescence in Arboreal Plants. *Biochemistry (Moscow)*. Volume 79, Issue 3, Pages 260-272.

DOI: 10.1134/S0006297914030122

<http://link.springer.com/article/10.1134%2FS0006297914030122>

SOLOVCHENKO A., LUKYANOV A., SOLOVCHENKO O., ET AL. (2014). Interactive effects of salinity, high light and nitrogen starvation on fatty acid and carotenoid profiles in *Nannochloropsis oceanica* CCALA 804. *European Journal of Lipid Science and Technology*. 2014. 116. 5. 635-644.

DOI:10.1002/ejlt.201300456

<http://onlinelibrary.wiley.com/doi/10.1002/ejlt.201300456/abstract>

SHTAIDA N., KHOZIN-GOLDBERG I., SOLOVCHENKO A., ET AL. (2014). Downregulation of a putative plastid PDC E1 α subunit impairs photosynthetic activity and triacylglycerol accumulation in nitrogen starved photoautotrophic *Chlamydomonas reinhardtii*. *Journal of Experimental Botany*. Volume 65.

DOI: 10.1093/jxb/eru374

<https://academic.oup.com/jxb/article/65/22/6563/2885037/Downregulation-of-a-putative-plastid-PDC-E1>

THWE A. A. AND KASEMSAP P. (2014). Quantification of OJIP Fluorescence Transient in Tomato Plants Under Acute Ozone Stress. *Kasetsart Journal: Natural Science*, Volume 48, Page 665 – 675.

http://kasetsartjournal.ku.ac.th/kuj_files/2015/A1501221529054531.pdf

AROCA R., RUIZ-LOZANO M. J., ZAMARREÑO A. M., ET AL. (2013). Arbuscular mycorrhizal symbiosis influences strigolactone production under salinity and alleviates salt stress in lettuce plants. *Journal of Plant Physiology*, Volume 170, Issue 1, Pages 47-55

DOI:10.1016/j.jplph.2012.08.020

<http://www.sciencedirect.com/science/article/pii/S0176161712004014>

GAJEWSKA E., DROBIK D., WIELANEK M. ET AL. (2013). Alleviation of nickel toxicity in wheat (*Triticum aestivum* L.) seedlings by selenium supplementation. *Biological Letters*. Volume 50, Issue 2, Pages 65–78.

DOI: 10.2478/biolet-2013-0008

<http://www.degruyter.com/view/j/biolet.2013.50.issue-2/biolet-2013-0008/biolet-2013-0008.xml>

PTUSHENKO V.V., PTUSHENKO E. A., SAMOILOVA O. P. ET AL. (2013). Chlorophyll fluorescence in the leaves of *Tradescantia* species of different ecological groups: Induction events at different intensities of actinic light. *Biosystems*. Volume 114, Issue 2, Pages 85–97.

DOI:10.1016/j.biosystems.2013.08.001

<http://www.sciencedirect.com/science/article/pii/S0303264713001810>

SOLOVCHENKO A., SOLOVCHENKO O., KHOZIN-GOLDBERG I., ET AL. (2013). Probing the effects of high-light stress on pigment and lipid metabolism in nitrogen-starving microalgae by measuring chlorophyll fluorescence transients: Studies with a $\Delta 5$ desaturase mutant of *Parietochloris incisa* (Chlorophyta, Trebouxiophyceae). *Algal Research*. Volume 2.

DOI 10.1016/j.algal.2013.01.01

VREDENBERG W. AND PAVLOVIČ A. (2012). Chlorophyll a fluorescence induction (Kautsky curve) in a Venus flytrap (*Dionaea muscipula*) leaf after mechanical trigger hair irritation. *Journal of Plant Physiology*. Volume 170, Pages 242-250.

DOI:10.1016/j.jplph.2012.09.009

<http://www.sciencedirect.com/science/article/pii/S017616171200404X>

CHYTYK, C. J., HUCL, P. J. AND GRAY, G. R. (2011). Leaf photosynthetic properties and biomass accumulation of selected western Canadian spring wheat cultivars. *Canadian Journal of Plant of Science*. Volume 91, Pages 305-314.

DOI: 10.4141/CJPS09163

<http://pubs.aic.ca/doi/abs/10.4141/CJPS09163>

COWLEY R. AND LUCKETT D. (2011) Chlorophyll fluorescence as a method to detect moisture-limiting stress in canola. 17th Australian Research Assembly on Brassicas (ARAB)

http://www.australianoilseeds.com/data/assets/pdf_file/0017/8306/S1-P1-Cowley.pdf

KOCUREK V., VONDRA M. AND SMUTNÝ, V. (2011). Efficacy of reduced doses of bentazone assessed by instruments based on measurement of chlorophyll fluorescence. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*. Volume 59, Pages 137-144

DOI: 10.11118/actaun201159010137

<http://acta.mendelu.cz/59/1/0137/>

KUVYKIN V., PTUSHENKO V. V., VERSHUBSKII A. V. ET AL. (2011). Regulation of electron transport in C₃ plant chloroplasts in situ and in silico: Short-term effects of atmospheric CO₂ and O₂. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, Volume 1807, Issue 3, Pages 336-347.

DOI:10.1016/j.bbabi.2010.12.012

<http://www.sciencedirect.com/science/article/pii/S0005272810007991>

LUCIŃSKI R., MISZTAŁ L. SAMARDAKIEWICZ S. ET AL. (2011). The thylakoid protease Deg2 is involved in stress-related degradation of the photosystem II light-harvesting protein Lhcb6 in *Arabidopsis thaliana* *New Phytologist*. Volume 192, Pages 74-86.

DOI: 10.1111/j.1469-8137.2011.03782.x.

<http://www.ncbi.nlm.nih.gov/pubmed/21668884>

RUIZ-SÁNCHEZ, M., ARMADA, E., MUÑOZ, Y., ET AL. (2011). Azospirillum and arbuscular mycorrhizal colonization enhance rice growth and physiological traits under well-watered and drought conditions. *Journal of Plant Physiology*. Volume 168, Issue 10, Pages 1031-1037.

DOI:10.1016/j.jplph.2010.12.019

<http://www.sciencedirect.com/science/article/pii/S0176161711000794>

SAMOILOVA O. P., PTUSHENKO V. V., KUVYKIN V. ET AL. (2011) Effects of light environment on the induction of chlorophyll fluorescence in leaves: A comparative study of *Tradescantia* species of different ecotypes. *Biosystems*. Volume 105, Issue 1, Pages 41-48.

DOI:10.1016/j.biosystems.2011.03.003

<http://www.sciencedirect.com/science/article/pii/S0303264711000566>

CESSNA S., DEMMIG-ADAMS B. AND ADAMS III W. W. (2010). Exploring Photosynthesis and Plant Stress Using Inexpensive Chlorophyll Fluorometers. *Journal of Natural Resources and Life Sciences Education*. Volume 39, Pages 22-30.

DOI: 10.4195/jnrlse.2009.0024u

http://www.researchgate.net/publication/227584624_Exploring_Photosynthesis_and_Plant_Stress_Using_Inexpensive_Chlorophyll_Fluorometers

FERNANDEZ-MARIN B., BECERRIL J. M. AND GARCIA PLAZAOLA J. I. (2010). Unravelling the roles of desiccation-induced xanthophyll cycle activity in darkness: A case study in *Lobaria pulmonaria*. *Planta*. Volume 231, Pages 1335-1342.

DOI: 10.1007/s00425-010-1129-6

<http://link.springer.com/article/10.1007/s00425-010-1129-6>

FROLEC J., ŘEBÍČEK J., LAZÁR D. ET AL. (2010). Impact of two different types of heat stress on chloroplast movement and fluorescence signal of tobacco leaves. *Plant Cell Reports*. Volume 29, Pages 705–714.

DOI: 10.1007/s00299-010-0856-2

<http://link.springer.com/article/10.1007%2Fs00299-010-0856-2>

PAVLOVIČ A., SLOVÁKOVÁ L., PANDOLFI C. ET AL. (2010). On the mechanism underlying photosynthetic limitation upon trigger hair irritation in the carnivorous plant Venus flytrap (*Dionaea muscipula* Ellis). *Journal of Experimental Botany*, Volume 62, Pages 1991–2000.

DOI: 10.1093/jxb/erq404

<http://jxb.oxfordjournals.org/content/early/2011/02/02/jxb.erq404.full>

RUIZ-SÁNCHEZ M., AROCA R., MUÑOZ Y., ET AL. (2010). The arbuscular mycorrhizal symbiosis enhances the photosynthetic efficiency and the antioxidative response of rice plants subjected to drought stress. *Journal of Plant Physiology*. Volume 167, Pages 862-869.

DOI: 10.1016/j.jplph.2010.01.018

<http://www.sciencedirect.com/science/article/pii/S0176161710000933>

HARDING S. A., JARVIE M. M., LINDROTH R. L. ET AL. (2009). A comparative analysis of phenylpropanoid metabolism, N utilization, and carbon partitioning in fast- and slow-growing *POPULUS* hybrid clones. *Journal of Experimental Botany*. Volume 60, Pages 3443-3452.

DOI:10.1093/jxb/erp180

<http://jxb.oxfordjournals.org/content/60/12/3443.full.pdf+html>

KUVYKIN I.V., VERSHUBSKII A.V., PRIKLONSKII V.I. ET AL. (2009). Computer simulation study of pH-dependent regulation of electron transport in chloroplasts. *Biophysics*. Volume 54, Pages 455-464.

DOI: 10.1134/S0006350909040101

<http://link.springer.com/article/10.1134%2FS0006350909040101>

MACEK P., MACKOVÁ J. AND DE BELLO F., (2009). Morphological and ecophysiological traits shaping altitudinal distribution of three *Polylepis* treeline species in the dry tropical Andes. *Acta Oecologica*, Volume 35, Pages 778–785.

DOI:10.1016/j.actao.2009.08.013

<http://www.sciencedirect.com/science/article/pii/S1146609X09001076>

ROSESCU M. R. AND ANDREI M. (2009). The study of photosystem II efficiency on selected synanthropic plant species. *Annals Food Science and Technology*. Volume 10, Pages 115-119.

http://www.afst.valahia.ro/docs/issues/13_Rosescu.pdf

BARTÁK, M (2008) *Biophysical Methods and Approaches to Monitor In-situ Lichen Responses to Environmental Extremes*. Coordination Action for Research Activities on life in Extreme Environments. Publication 2.

<http://carex->

[eu.ameos.net/fileadmin/user/upload/Workshops/Project_Forum/Proceedings_ModEco_Final.pdf](http://carex-eu.ameos.net/fileadmin/user/upload/Workshops/Project_Forum/Proceedings_ModEco_Final.pdf)

KLEM K. AND BAJEROVA, E., (2008). *Adjustment of herbicide dose in sugar beet based on non-invasive chlorophyll fluorescence measurements. Agricultural And Biosystems Engineering For A Sustainable World: National Conference On Agricultural Engineering, Hersonissos, Crete, Greece, Pages 23-25.*

<http://www.cabdirect.org/abstracts/20083323643.html>

WOO N. S., BADGER M. R. AND POGSON B. J. (2008) *A rapid, non-invasive procedure for quantitative assessment of drought survival using chlorophyll fluorescence Plant Methods, Volume 4, Issue 27, Pages 1-14.*

DOI:10.1186/1746-4811-4-27

<http://www.plantmethods.com/content/4/1/27>

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